

Enzymatic Assay of α -Fucose Dehydrogenase

Introduction

The monosaccharide α -fucose (6-deoxy-L-galactose) is a component of many N- and O-linked glycans and glycolipids and is produced by microorganisms, plants, and animals. It is a common terminal modification of glycan structures and has a role in many different processes. In addition, a change in α -fucose metabolism has been linked to different diseases including breast cancer, ovarian cancer, colorectal adenocarcinoma, leukemia, brain tumors, cirrhosis, meningitis, tuberculosis, and cardiovascular disorders. One way to study α -Fucose is to measure its metabolism by α -fucose dehydrogenase, the main enzyme responsible for α -fucose catabolism. Here, we outline a procedure in which the oxidation of α -fucose by α -fucose dehydrogenase is measured by UV spectroscopy and a reaction rate is established. This can be used in other applications to determine rates of reaction for other enzymes that utilize α -fucose.

Materials

- Tris (GoldBio Catalog # [T-400](#))
- Imidazole (GoldBio Catalog # [I-902](#))
- Acetate HCl buffer
- 5M HCl
- L-Fucose (GoldBio Catalog # [F-260](#))
- Molecular biology grade water
- Ice
- β -NADP
- Spectrophotometer with a path length of 1 cm

Preparation of Buffers and Solutions:

- Prepare these reagents with the final concentrations listed. The buffer should have a pH of 9.5 at a temperature of 37°C. Adjust the pH with 5M HCl if necessary.
 - a. 120mM Tris
 - b. 120mM Imidazole
 - c. 100mM Acetate HCl Buffer

For the α -Fucose Solution:

- Prepare a fresh solution with a final concentration of 150mM in molecular biology grade water.

- Solution should be used within an hour of preparation and should be kept on ice at all times.

For the β -NADP Solution:

- Prepare a solution of β -Nicotinamide Adenine Dinucleotide Phosphate (β -NADP) with a final concentration of 15mM in cold molecular biology grade water.

For the α -Fucose Dehydrogenase Enzyme Solution:

- Prepare a solution of isolated α -Fucose Dehydrogenase in cold molecular biology grade water immediately before use.

Method

1. Into a cuvette prepare the following test and blank samples (see Table 1). Invert to mix and allow to equilibrate to 37°C. Monitor the $A_{340\text{ nm}}$ until constant.

Table 1. Preparation of test and blank samples.

Reagent	Test (mL)	Blank (mL)
Buffer	2.50	2.50
α -Fucose	0.20	N/A
β -NADP	0.20	0.20
Molecular Biology Grade Water	N/A	0.20

2. Add 0.10 mL of the enzyme solution to the test and blank, quickly invert to mix and measure the change in $A_{340\text{ nm}}$ /min for 15 minutes.

Note: In the presence of NADP, 1 unit of α -Fucose Dehydrogenase will typically oxidize 1.0 μ M of α -Fucose into α -Fucono-1,5-lactone per minute (at a pH of 9.5 and 37°C).

Calculations

$$\frac{\text{Units}}{\text{mL}} \text{ enzyme} = \frac{(\Delta A_{340\text{ nm}}/\text{min}_{\text{Test}} - \Delta A_{340\text{ nm}}/\text{min}_{\text{Blank}})(3)(df)}{(6.22)(0.1)}$$

3 = Volume (in ml) of assay

Df = dilution factor (if necessary)

6.22 = Millimolar extinction coefficient of β -NADPH at 340 nm

0.1 = Volume (in ml) of enzyme solution used

Associated Products

- [Imidazole \(GoldBio Catalog # I-902\)](#)
- [L-Fucose \(GoldBio Catalog # F-260\)](#)
- [Tris \(GoldBio Catalog # T-400\)](#)

References

Becker, D. J. and Lowe, J. B. (2003). Fucose: Biosynthesis and biological function in mammals. *Glycobiology*, 13(7). Doi:10.1093/glycob/cwg054.

Horiuchi, T., Suzuki, T., Hiruma, M. and Saito, N. (1989) Purification and Characterization of α -Fucose (α -Galactose) Dehydrogenase from *Pseudomonas sp.* No. 1143. *Agricultural and Biological Chemistry*, 53, 1493-1501. Doi: 10.1080/00021369.1989.10869508.